

PRODIGIOSIN-LIKE PIGMENTS FROM
ACTINOMADURA (NOCARDIA) PELLETIERI

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Two prodigiosin-like pigments from *Actinomadura pelletieri* were shown to be undecylprodiginine (III) and methylcyclodecylprodiginine (VI). The antimicrobial activity of four prodiginine pigments is given.

Prodigiosin¹⁾ (I) is the bright red pigment of *Serratia marcescens* and was probably responsible for many medieval "miracles" involving the appearance of blood stains on the Holy Host²⁾. Two red pigments with the methoxytripyrrole nucleus of I (prodiginine) have been isolated from *Streptomyces longisporus ruber*. They are undecylprodigiosin³⁾ (undecylprodiginine) (III) and metacycloprodigiosin (IV)⁴⁾. Recently we reported nonylprodiginine (II)⁵⁾ and cyclononylprodiginine (V)⁶⁾ to be produced by red strains of *Actinomadura*⁷⁾ (*Nocardia madurae*).

We have detected, by thin-layer chromatography (TLC) and visible absorption spectroscopy, similar pigments in 16 strains of the closely related organism, *A. pelletieri*, which is normally red. The slower moving TLC spot was shown, in the case of strain 687, to be undecylprodiginine (III) by comparison with an authentic sample. Although II and III have identical visible absorption spectra and similar TLC behavior, they can be readily distinguished on a 0.1 mg scale by their partition coefficients. The K's for II and III in C₆H₆ - EtOH - H₂O - HOAc (15:10:5:1) were 1.5 and 2.5 respectively.

The pigment corresponding to the faster moving TLC spot was obtained in quantity from strain 610. A comparison of it with V showed some similarities. The visible absorption maximum was 539 m μ in acidic EtOH, 549 in hexane. In the mass spectrum, the base peak was the molecular ion at 391, M⁺⁺ at 195.5 was also strong. The nmr spectrum showed two singlets at 6.7 and 5.8 δ which were assigned to hydrogens at carbons 5 and 7 as well as bands characteristic of two AB systems each containing one " α " and one " β " type of pyrrole proton⁸⁾. Thus (as has been previously explained in greater detail⁶⁾) the alkyl chain must be attached at positions 2 and 10.

However, in contrast to V, the nmr spectrum of VI showed an area corresponding to 2 hydrogens, not 4, at the characteristic location for $-\text{CH}_2-\text{Ar}$, 2.9 δ . Furthermore, the mass spectrum (see Fig. 2) of V shows that fragmentation ions with 1 and 3 $-\text{CH}_2-$ units derived from the side chain predominated over those with 0, 2, 4, 5 or more $-\text{CH}_2-$ units. In contrast, from VI, ions with 2 and 4 $-\text{CH}_2-$ units predominated. This

suggested that one carbon α to the aromatic portion of the molecule had a methyl branch on it. If this were the case then one might expect VI to be optically active. The circular dichroism of V and VI were measured in EtOH. The tracing for V was flat while that for VI showed a small but definite peak at 530 m μ indicating optical activity not due to impurities.

Previously, we had investigated II and V by oxidation with dilute potassium permanganate in aqueous pyridine⁶). The acids obtained were methylated and analyzed by gas chromatography (GC). Pigment II gave a 1:4 mixture of methyl nonate and methyl decanoate; V furnished a 2:7:5 mixture of the dimethyl esters of azelaic, sebacic, and undecanedioic acids. All were identified by comparison with authentic samples and the structure of dimethyl sebacate was verified by its mass spectrum. Thus, in this oxidation, one obtains a mixture of acids derived from the aliphatic part of the molecule. Oxidative cleavage may occur α to the pyrrole ring (methyl nonate from II) or in such a way that the product includes one of the ring carbons (methyl decanoate from II). For V, one would expect 3 different dicarboxylic acids with sebacic acid predominating, since it could be formed in two different ways.

However, VI did not furnish the dimethyl esters of undecanedioic, dodecanedioic and tridecanedioic acids as would have been expected from an unbranched eleven carbon aliphatic chain. Instead, GC revealed 5 main peaks whose retention times are given in Fig. 3 compared to those for sebacic, undecanedioic and dodecanedioic dimethyl esters. For a homologous series run under isothermal conditions, the log of

Fig. 1. Structures of the prodiginine pigments

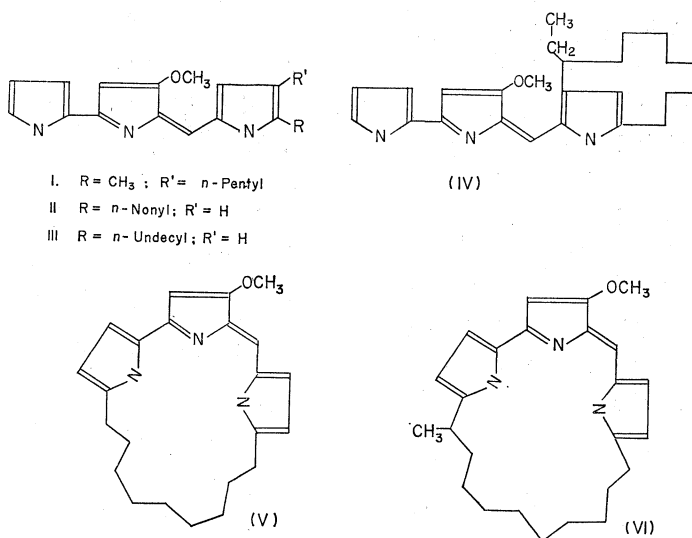
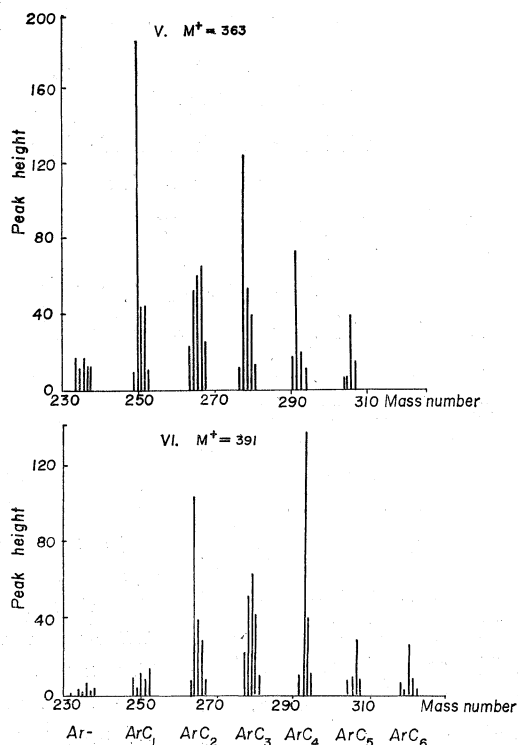


Fig. 2. Portions of the mass spectra of V and VI



the retention time versus carbon number is a straight line (except for the lowest members of the series). Series of esters having the same type of characteristic groupings give parallel straight lines⁹). Thus, Fig. 2 shows that B and D are homologs, as well as E and G, but they are not members of the same series. Material from peaks B and D gave a positive microtest for carbonyl with 2,4-dinitrophenylhydrazine¹⁰). Furthermore, B was identical with an authentic sample of methyl 10-oxoundecanoate in GC retention time and TLC behavior of the dinitrophenylhydrazone derivatives. Thus B and D are methyl 10-oxoundecanoate and methyl 11-oxododecanoate where oxidative cleavage at the branched end has occurred α to the pyrrole ring. The other pair of products, in which at the branched end one carbon from the pyrrole ring is included, would be α -methylundecanedioic and α -methyl dodecanedioic acids. A direct comparison with an authentic sample was not possible but the retention times of E and G are consistent with these structures since branched chain fatty esters have shorter retention times than the corresponding unbranched ones⁹).

The retention time of peak F was identical with that of dimethyl dodecanedioate, however its distinct "mushroom" odor suggested that it was some other type of degradation product since the diesters are odorless under our GC conditions. One might imagine that VI was a mixture of pigments containing enough of one with an unbranched ten carbon chain to furnish dimethyl dodecanedioate on oxidation. However, this was disproved by the complete absence of any M^{++} at 188.5 in the mass spectrum of VI.

Although most strains of *A. pelletieri* gave VI; strain 757 furnished II and V. Six red strains of *A. madurae* were available for study. The majority furnished II and V, 554 produced only III, shown by its partition coefficient compared to those of II and III. Strain 507 gave a pigment whose partition coefficient was intermediate between those of II and III. The mass spectrum demonstrated that it was a mixture of II and III (M^+ at 393 and 365) not a pigment with a 10 carbon side chain. In all cases strain identification had been made previously¹¹). The presence of C-9 and C-11 side chains but not C-10; as well as coproduction of II and III

Fig. 3. The retention time of the methyl esters obtained from VI and of the 3 standard dimethyl esters

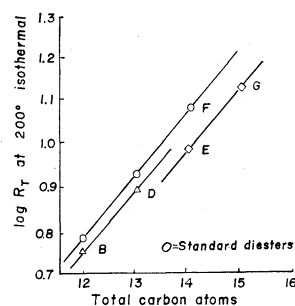


Table 1. Antimicrobial activity of proginine pigments

Organism	Minimum inhibitory concentration ($\mu\text{g}/\text{ml}$ agar)			
	I	II	V	VI
<i>Staphylococcus aureus</i> 15	10, 5*	25	25	25
<i>Mycobacterium smegmatis</i> 3	10, 5*	50*, 25*	25	25
<i>Nocardia asteroides</i> 3318	25, 10*	50*, 25*	50*, 25*	25*
<i>Actinomadura madurae</i> R-28	10	50, 25*	25	25
<i>Streptomyces somaliensis</i> 395	10	25	25, 10*	10
<i>M. rhodochrous</i> 271	10, 5*	50, 25*	25*	25
<i>Trichophyton mentagrophytes</i> 171	10, 5*	50*	25*	25*
<i>Ceratostomella ulmi</i> 185	50, 25*	(-)**	(-)	(-)
<i>Saccharomyces cerevisiae</i> 216	25*	(-)	(-)	(-)
<i>Prototheca zopfii</i> 328	25, 10*	(-)	(-)	(-)
<i>Pacilomyces varioti</i> 3	25*	(-)	(-)	(-)
<i>Aspergillus niger</i> 13	25*	(-)	(-)	(-)

* Partial inhibition

** (-)=Not active

by strain 507 suggests that the side chains are biosynthesized from acetate units by chain extension as in fatty acids. Then, at some point before the extended chain is fully reduced (for example: at the R-CH=CH-COOH stage), cyclization may occur. The methyl branch in VI is considered to be α to ring A not ring C on the basis of these biogenetic suggestions which are for the actinomycetic prodiginine pigments only. The terminal stage in the biosynthesis of prodigiosin has been shown to involve coupling of 4-methoxy-2,2'-bipyrrole-5-carboxaldehyde and 2-methyl-3-ampyrrole¹¹.

The antimicrobial activity of pigments I, II, V and VI is shown in Table 1. Prodigiosin (I), showed modest activity against some gram positive bacteria, actinomycetes and fungi. The others had a similar but weaker activity.

Experimental

The production, isolation and purification of the prodiginine pigments has been described previously⁹.

Authentic 10-oxoundecanoic acid was prepared from 10-undecynoic acid using the method of SISO¹²) and esterified with BCl₃-MeOH.

Permanganate Oxidation of VI. To 18 mg of VI perchlorate salt in 4 ml of pyridine was added 5 ml of NaIO₄-KMnO₄ solution (1 g of NaIO₄+0.1 g KMnO₄/100 ml distilled water) and 25 mg of KMnO₄. The mixture was shaken at 28°C additional portions of KMnO₄ were added after 4 hours (10 mg), 4 hours (10 mg) and overnight (10 mg). Control flasks with pyridine only or with pyridine and 10-oxoundecanoic acid to which the NaIO₄-KMnO₄ solution had been added were not decolorized during several days at 28°C. Four hours after the last addition of KMnO₄, the reaction mixture was centrifuged; the MnO₂ was stirred with very dilute aqueous Na₂CO₃, centrifuged again and the combined supernatants decolorized by warming with a few drops of EtOH. After centrifugation this supernatant was acidified with H₂SO₄ and extracted 5 times with CHCl₃. The residue from the CHCl₃ solution (8 mg) was esterified with BCl₃-MeOH and analyzed by gas chromatography on an F and M model 700 dual column machine equipped with a thermal conductivity detector and a 180×0.65 cm column of 10% SE-30 on Diatoport 60~80 mesh using He flow at 50 ml/min. The 5 essentially equal-sized peaks had retention times between 29 and 36 minutes when the column temperature was started at 80°C and raised 4°C/min. Peak areas indicated a total yield of about 1.5 mg. There were no peaks in the methyl decanoate range (about 20 minutes under these conditions). Low-boiling esters such as dimethyl oxalate would have been lost during evaporation of solvent.

Antimicrobial Assays. Weighed samples of the pure perchlorate salts were dissolved in CHCl₃ and transformed into the free base form by shaking with aqueous NH₃. The CHCl₃ solutions were evaporated at room temperature under a stream of CO₂ and the vacuum-dried residues dissolved in distilled dimethylsulfoxide. Assays were carried out using the agar-streak dilution method¹³) on nutrient agar+0.5% NaCl and 1.0% glucose. Each assay was run twice and compared to a solvent control. All were incubated at 28°C. Final readings were made at 24 hours for bacteria, 48 hours for mycobacteria and some filamentous fungi, 5 and 7 days for *Ceratostomella ulmi*, *Trichophyton mentagrophytes* and all actinomycetes.

None of the compounds tested had any activity against *Escherichia coli* 54, *Proteus vulgaris* 73, *Candida albicans* 204, *Trichoderma koningi* 55 or *Penicillium notatum* 40. Prodigiosin was inactive against *Pseudomonas aeruginosa* 77. The concentrations used for I were 50, 25, 10, 5 and 1 μ g/ml of agar; the other three were tested at 50, 25 and 1 μ g/ml. Dilutions were in dimethylsulfoxide.

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